

MoFlo™ Viability and Function with High-Speed Sorting

Introduction

Sustained cell sorting at speeds in excess of 50,000 events per second is routine when using the MoFlo high-performance cell sorter. This powerful instrument gives academic and clinical researchers the capability to perform laboratory protocols that were previously unrealistic or impractical due to the length of sort time required when using other instrumentation at slower sort rates. In addition to purity and recovery, MoFlo utilizes cell viability and function as endpoint measurements to benchmark high-speed sort performance.

High-Speed Sorting Applications

High-speed sorting enables a wide range of novel applications, particularly those related to isolation of rare biological events.1 Traditionally in this arena, scientists have relied on conventional flow cytometers in combination with technologies such as magnetic particle separation, centrifugal elution, densitygradient separation and complement-mediated lysis, to isolate specific cell populations. However, these technologies are time consuming and involve additional manipulation of the cell preparation that may lead to undesirable changes in cell function and/or cellular activation. High-speed sorting permits the use of a single purification step, delivering the cells of interest into culture or into the transfer recipient at >99% purity in the shortest possible time.

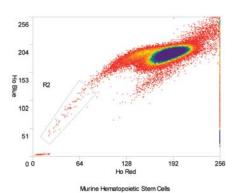


Figure 1

Murine Hematopoietic Stem Cells. Murine bone marrow is incubated with the nucleic acid dye Hoechst 33342, then analyzed using a MoFlo cell sorter. The Hoechst dye is excited using a 351 nm UV laser and fluorescent emission is detected both in the blue region, using a 450/20 bandpass filter, and in the red region, using a 675 eFLP filter. The progenitor cells of interest are identified by their characteristic position to the left of the bulk cell population. Acquisition and analysis were done using Summit software. Data courtesy of the University of Colorado Health Sciences Center.

With the MoFlo, operators can define multiple cell populations, based on selectable parameters and simultaneously distribute those populations – even extremely rare ones - in up to four separate temperature-controlled receptacles. MoFlo's patented electronics process events at such high speeds that cells of interest can be retrieved from the original sample with a single pass through the flow cytometer, without prior enrichment steps. Interest in the use of high-speed sorting is growing for applications related to stem cells, dendritic cells, rare thymus-borne T cell precursors and genetic transfectants, which may make up as little as 0.01% of a given cell population. Operators can now routinely perform sorts on these populations in much less time than it takes to setup and run a magnetic column.

Instrument Design

The challenge of high-speed sorting is to find a way of rapidly processing events while maintaining the ability to detect even dim fluorescence with high sensitivity. By concentrating the cell sample and running at high pressures (typically 60 psi or greater), the diameter of the core stream is maintained, ensuring high fluorescent measurement precision. Furthermore, the pressure, combined with size of the nozzle tip, determines the frequency at which droplets may be stably formed. A higher droplet frequency correlates to increased yield, as the likelihood of having multiple cells within a single drop decreases. Therefore, fewer droplets are discarded due to event coincidence. While legacy-

type sorters typically create 10,000-30,000 droplets per second, the MoFlo creates 90,000-200,000 droplets per second, significantly increasing the yield of every sort without sacrificing purity. Moving cells at these speeds requires careful design of the fluidics system, not only to maintain laminar flow, but also to ensure that sorted cells remain viable and fully functional after isolation.

Proven Performance

The MoFlo uses a patented nozzle design to reduce turbulence and minimize the effects of acceleration on each cell. This design has been extensively validated in peer-reviewed publications. Scientists worldwide routinely use the MoFlo to identify and segregate a wide variety of cell types including T cells, B cells, NK cells, dendritic cells, hematopoietic (Figure 1) and neural stem cells. Following sorting, these cells are fully functional and capable of cytokine production, antigen presentation, antibody production, activation, target-cell binding, posttransplantation engraftment and long-term culture.2-32

References

- 1. Ashcroft RG and Lopez PA. Commercial high-speed machines open new opportunities in high-throughput flow cytometry. Journal of Immunological Methods 2000; 243:13-24.
- 2. Arnold LW et al. Identification of a precursor to phosphatidyl choline-specific B-1 cells suggesting that B-1 cells differentiate from splenic conventional B cell in vivo: Cyclosporin a blocks differentiation to B-1. J Immunol 2000; 164: 2924-2930.
- 3. Avitahl N et al Ikaros sets thresholds for T cell activation and regulates chromosome propagation. Immunity 1999; 10: 333-343.
- 4. Batten Ml. BAFF mediates survival of peripheral immature B lymphocytes. J Exp Med 2000; 192(10): 1453.
- 5. Chatterjee S et al. Transduction of primitive human marrow and cord blood-derived hematopoietic progenitor cells with adeno-associated virus vectors. Blood 1999; 93(6): 1882-
- 6. Cipriani B et al. Activation of C-C B-chemokines in human peripheral blood T cells by isopentenyl pyrophosphate and regulation by cytokines. Blood 2000; 95(1): 39.
- 7. Clark LB et al. Cellular and molecular characterization of the scurfy mouse mutant. J Immunol 1999; 162: 2546-2554.
- 8. Dahl AM et al. Expression of BCL-XL restores cell survival, but not proliferation and effector differentiation, in CD28-deficient T lymphocytes. J Exp Med 2000; 191(12): 2031-2037.
- 9. Gartner F et al. 2000. Antigen-independent appearance of Recombination Activating Gene (RAG)-positive bone marrow B cells in the spleens of immunized mice. J Exp Med 2000; 192(12): 1745.
- 10. Ho IC et al. c-maf promotes T helper cell type 2 (Th2) and attenuates the Th1 differentiation by both interleukin 4-dependent and -independent mechanisms. J Exp Med 1998; 188(10): 1859.
- 11. Hoshino T et al. IL-18 is a potent coinducer of IL-13 in NK and T cells: a new potential role for IL-18 in modulating the immune response. J Immunol 1999; 162: 5070-5077.
- 12. Hoshino T et al. IL-13 production by NK cells: IL-13-producing NK and T cells are present in vivo in the absence of IFN-a. J Immunol 1999; 162: 51-59.
- 13. Hulspas R et al. Characterization of neurosphere cell phenotypes by flow cytometry. Cytometry 2000; 40: 245-250.
- 14. Kim JI et al. The transcription factor c-Maf controls the production of interleukin-4 but not other Th2 cytokines. Immunity 1999; 10: 745-751.

- 15. Kouro T et al. Characteristics of early murine B-lymphocyte precursors and their direct sensitivity to negative regulators. Blood 2001; 97(9): 2708-2715.
- 16. Kubota H, Reid LM. Clonogenic hepatoblasts, common precursors for hepatocytic and biliary lineages, are lacking classical major histocompatibility complex class I antigen. PNAS 2000; 97(22): 12132.
- 17. Le Bon A et al. Type I interferons potently enhance humoral immunity and can promote isotype switching by stimulating dendritic cells in vivo. Immunity 2001; 14(4): 461-470.
- 18. Li Y et al. Mast cells/basophils in the peripheral blood of allergic individuals who are HIV-1 susceptible due to their surface expression of CD4 and the chemokine receptors CCR3, CCR5, and CXCR4. Blood 2001; 97(11): 3483-3490.
- 19. Mason LH et al. Interaction of Ly-49D+ NK cells with H-2Dd target cells leads to Dap-12 phosphorylation and IFN-a secretion. J Immunol 2000; 164: 603-611.
- 20. Nakagawa TY et al. Impaired invariant chain degradation and antigen presentation and diminished collagen-induced arthritis in Cathepsin S null mice. Immunity 1999; 10: 207-217.
- 21. Ortaldo JR et al. Structure/function relationship of activating Ly-49D and inhibitory Ly-49G2 NK receptors. J Immunol 1999; 163: 5269-5277.
- 22. Ouyang W et al. Stat6-independent GATA-3 autoactivation directs IL-4-independent Th2 development and commitment. Immunity 2000; 12: 27-37.
- 23. Pestano GA et al. Inactivation of misselected CD8 T cells by CD8 gene methylation and cell death. Science 1999; 284:
- 24. Poggi A et al. IL-12-mediated NKRP1A up-regulation and consequent enhancement of endothelial transmigration of $V\alpha 2+TCR\delta\alpha+T$ lymphocytes from healthy donors and multiple sclerosis patients. J Immunol 1999; 162: 4349-4354.
- 25. Shih C et al. Long-term ex vivo maintenance and expansion of transplantable human hematopoietic stem cells. Blood 1999; 94(5): 1623-1636.
- 26. Spyridonidis A et al. Purging of mammary carcinoma cells during ex vivo culture of CD34+ hematopoietic progenitor cells with recombinant immunotoxins. Blood 1998; 91(5): 1820-1827.
- 27. Stewart FM et al. Lymphohematopoietic engraftment in minimally myeloablated hosts. Blood 1998; 91(10): 3681-
- 28. Stockschlader M et al. Expansion and fibronectin-enhanced retroviral transduction of primary human T lymphocytes for adoptive immunotherapy. Journal of Hematotherapy and Stem Cell Research 1999; 8(4): 401-410.
- 29. Teague TK et al. Activation-induced inhibition of interleukin 6mediated T cell survival and signal transducer and activator of transcription 1 signaling. J Exp Med 2000; 191(6): 915-925.
- 30. Usherwood EJ et al. Functionally heterogeneous CD8+ T cell memory is induced by Sendai virus infection of mice. J Virol 1999; 73(9): 7278-7286.
- 31. Van Rij RP et al. Differential coreceptor expression allows for independent evolution of non-syncytium-inducing and syncytium-inducing HIV-1. J Clin Investigation 2000; 106(8): 1039.
- 32. Wilson SB et al. Multiple differences in gene expression in regulatory V 24J Q T cells from identical twins discordant for type I diabetes. PNAS 2000; 97(13) 7411.

For Research Use Only. Not for use in Diagnostic Procedures.



UK, High Wycombe (44) 01494 441181 USA, Fullerton, CA (1) 800 742 2345